



## **2015 SAMPLING AND ANALYSIS WORKPLAN FOR BASELINE MONITORING OF THE SOUTH FORK BLUE RIVER**

PREPARED BY

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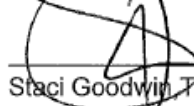
## 2015 Sampling and Analysis Workplan for Baseline Monitoring of the South Fork Blue River

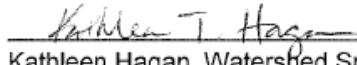
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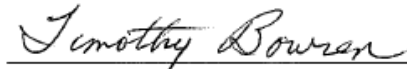
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### Reviews and Approvals

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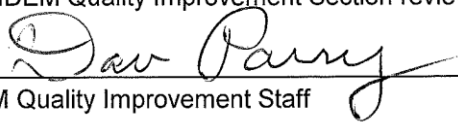
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The IDEM Quality Improvement Section reviewed and approves this Sampling and Analysis Workplan.

 Date 1-26-2014  
IDEM Quality Improvement Staff

### **Work Plan versus QAPP:**

This Sampling and Analysis Work Plan is an extension of the existing Watershed Assessment and Planning Branch, October 2004 “*Quality Assurance Project Plan (QAPP) for Indiana Surface Water Quality Monitoring and Total Maximum Daily Load (TMDL) Program*” and serves as a link to the existing QAPP as well as an independent QAPP of the project. Per the U.S. EPA QAPP guidance, this Work Plan establishes criteria and specifications pertaining to a specific water quality monitoring project that are usually described in the following four groups (phases) or sections as QAPP elements:

#### **Phase A. Project Management/Planning**

The plan documents project history and objectives, and establishes Data Quality Objectives (DQOs).

#### **Phase B. Measurement/Data Acquisition**

The plan describes sampling procedures, analytical methods, sample and data acquisition requirements, and the quality control measures specific to the project.

#### **Phase C. Assessment/Oversight**

The plan identifies the key elements of external and internal checks, audits, peer reviews, Data Quality Assessments (DQAs), and the preparation of Quality Assurance/Quality Control (QA/QC) Review Reports for management.

#### **Phase D. Data Validation and Usability**

The plan describes data handling and associated QA/QC activities, including QA/QC Review Reports.

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## LIST OF ACRONYMS

AAC:	Acute Aquatic Criterion
ADC:	Acoustic Doppler Current
ADP:	Acoustic Doppler Profiler
ADV:	Acoustic Doppler Velocimeter
AIMS:	Assessment Information Management System
CAC:	Chronic Aquatic Criteria
CALM:	Consolidated Assessment Listing Methodology
CCC:	Criterion Continuous Concentration
CDL:	Crop Data Layer
CFR:	Code of Federal Regulations
CFU:	Colony Forming Units
CLP:	Contract Laboratory Program
COD:	Chemical Oxygen Demand
CPR:	Cardio-Pulmonary Resuscitation
CRQL:	Contract Required Quantification Limit
DO:	Dissolved Oxygen
DQA:	Data Quality Assessment
DQO:	Data Quality Objectives
E. coli:	Escherichia coli
EPA:	Environmental Protection Agency
GPS:	Global Positioning System
HUC:	Hydrologic Unit Code
IAC:	Indiana Administrative Code
IBC:	Impaired Biotic Community
IBI:	Index of Biotic Integrity
IDEM:	Indiana Department of Environmental Management
MDL:	Method Detection Limit
µS/cm	Micro Siemens per Centimeter
mg/L:	Milligram per liter
MHAB:	Multi-habitat
mL:	Milliliter
MPN:	Most Probable Number
MS/MSD:	Matrix Spike/Matrix Spike Duplicate
NTU:	Nephelometric Turbidity Unit(s)
OWQ:	Office of Water Quality
PFD:	Personal Floatation Device
PPE:	Personal Protective Equipment
QA/QC:	Quality Assurance/Quality Control
QAC:	Quality Assurance Coordinator
QAM:	Quality Assurance Manager

QAO:	Quality Assurance Officer
QAPP:	Quality Assurance Project Plan
QHEI:	Qualitative Habitat Evaluation Index
RFP:	Request for Proposals
RL:	Reporting Limit
RPD:	Relative Percent Difference
S.U.:	Standard Units
SM:	Standard Method
SOP:	Standard Operating Procedures
TDS:	Total Dissolved Solids
TKN:	Total Kjeldahl Nitrogen
TMDL:	Total Maximum Daily Load
TOC:	Total Organic Carbon
TP:	Total Phosphorus
TS:	Total Solids
TSS:	Total Suspended Solids
U.S.:	United States
USDA:	United States Department of Agriculture
WAPB:	Watershed Assessment and Planning Branch

## ***Definitions:***

Elutriate	To purify, separate, or remove lighter or finer particles by washing, decanting, and settling.
Geometric site	Sampling site chosen according to its drainage area within a watershed.
Fifteen (15) Minute Pick	A component of the IDEM multihabitat macroinvertebrate sampling method in which the one minute kick sample and fifty meter sweep sample collected at a site are combined, elutriated, with macroinvertebrates removed from the resulting sample for 15 minutes while in the field.

Fifty (50) Meter Sweep	A component of the IDEM multihabitat macroinvertebrate sampling method in which approximately 50 meters (50m) of shoreline habitat in a stream or river is sampled with a standard 500 micrometer (500 $\mu$ m) mesh width D-frame dipnet by taking 20-25 individual “jab” or “sweep” samples, which are then composited.
One (1) minute kick sample	A stationary sampling accomplished using a box shaped net comprised of canvas bottom and/or sides and 504 $\mu$ nylon mesh back. The designated area is sampled for one minute.
Pour point	The outlet of a subwatershed or the common point where all the water flows out of any given subwatershed.
Targeted site	A sampling site intentionally selected based on specific monitoring objectives or decisions to be made.

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## Baseline Monitoring Program Objective

Baseline Monitoring uses an intensive targeted watershed design that characterizes the current condition of an individual watershed. This type of monitoring provides valuable data for the purposes of assessment, Total Maximum Daily Load (TMDL) development, watershed planning, and allows for future comparisons to evaluate changes in the water quality within the watershed(s) studied. Selecting a spatial monitoring design with sufficient sampling density to accurately characterize water quality conditions is a critical step in the process of developing an adequate local scale watershed study.

The Indiana Department Environmental Management (IDEM) has selected the South Fork Blue River Watershed (see Figure 1, Table 1) for a water quality baseline study. Sample sites were chosen using a modified geometric site selection process as well as targeted site selection in order to get the necessary spatial representation of the entire study area. Sites within this watershed were selected based on a geometric progression of drainage areas starting with the area at the mouth of the main stem stream and working upstream through the tributaries to the headwaters. Monitoring sites were then located to the nearest bridge. A more complete description of the geometric site selection process is included as Attachment 1. Sample sites were also chosen at the nearest bridge to the pour point (the lowest point in the basin through which all water flows) of each 12 digit Hydrologic Unit Code (HUC) in the watershed, or chosen to characterize sources for TMDL development.

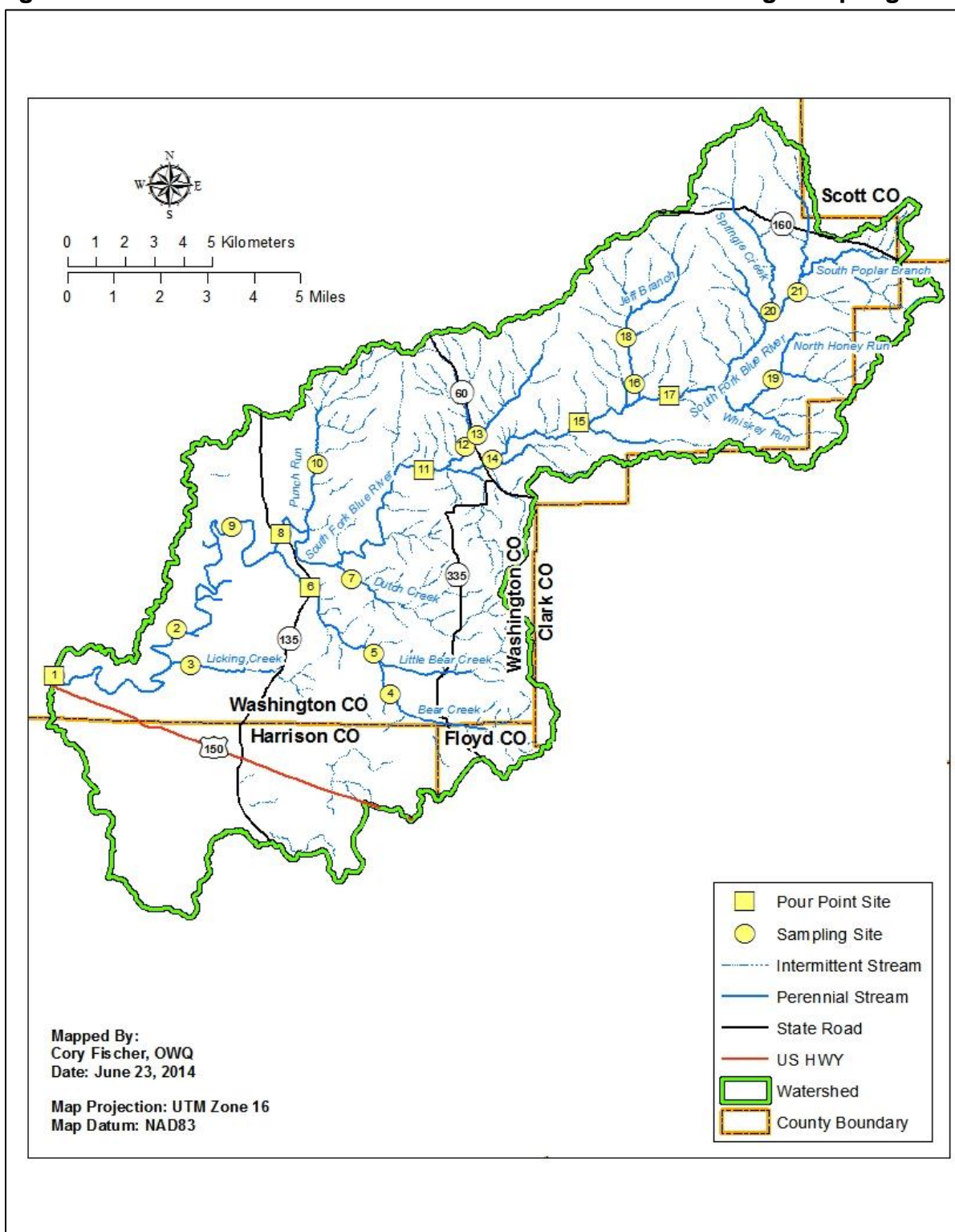
It is anticipated that the water quality data collected through this monitoring effort will provide the information needed to characterize the watershed for the TMDL program and local water quality managers, identify sources of impairment, designate critical areas, and enable users to make valid and informed watershed decisions. This project, by design, will also add new stream reaches for assessment of aquatic life and recreational use support and will allow for future comparisons to evaluate changes in water quality.

The draft 2014 303(d) list submitted to the U.S. EPA (IDEM 2014a) details impairments of approximately 38 miles of the South Fork Blue River Watershed in the following ways:

- Category 5(a): Impaired Biotic Community (IBC), 2.00 miles
- Category 5(a): *Escherichia coli* (*E. coli*), 31.25 miles
- Category 5(a): Dissolved Oxygen (DO), 4.38 miles

Assessment data in this watershed have been collected by IDEM from multiple programs and projects.

**Figure 1. South Fork Blue River Watershed Baseline Monitoring Sampling Area<sup>1</sup>**



<sup>1</sup> Map site numbers refer to last two digits of site number from Table 1; e.g., 15T-010 is site 10 on map

**Table 1. Sampling Locations for Baseline Monitoring of the South Fork Blue River<sup>3</sup>**

Site #	AIMS Site #	Stream Name	Location	County	Latitude	Longitude
15T-001	OBS130-0002	South Fork Blue River	Fredericksburg Road	Washington	38.43444639	-86.18341847
15T-002	OBS-06-0016	South Fork Blue River	Palmyra Rd	Washington	38.44858611	-86.13521301
15T-003	OBS-06-0015	Licking Creek	Palmyra Rd	Washington	38.43741894	-86.12961261
15T-004	OBS-06-0021	Bear Creek	Wetzel Rd	Washington	38.42770392	-86.05099359
15T-005	OBS-06-0014	Bear Creek	Martinsburg Fire Rd	Washington	38.44043334	-86.05749129
15T-006	OBS-06-0013	Bear Creek	SR 135	Washington	38.46143258	-86.08194909
15T-007	OBS-06-0007	Dutch Creek	Dutch Creek Rd	Washington	38.46393356	-86.06601976
15T-008	OBS-06-0008	South Fork Blue River	SR 135	Washington	38.47850981	-86.09370001
15T-009	OBS-06-0020	South Fork Blue River	Big Springs Rd	Washington	38.4805845	-86.1130785
15T-010	OBS-06-0009	Punch Run	Shorts Corner Rd	Washington	38.50003445	-86.07901664
15T-011	OBS-06-0004	South Fork Blue River	Martinsburg Rd	Washington	38.49812761	-86.03677311
15T-012	OBS-06-0006	Tributary of South Fork Blue River	Shorts Corner Rd	Washington	38.50531258	-86.02005328
15T-013	OBS-06-0012	Tributary of South Fork Blue River	Mahuron Rd	Washington	38.50859	-86.015463
15T-014	OBS-06-0018	South Fork Blue River	Main St	Washington	38.50095172	-86.0096978
15T-015	OBS-06-0022	South Fork Blue River	Lockenour Rd	Washington	38.51238886	-85.9751927
15T-016	OBS-06-0003	Jeff Branch	E Blue River Rd	Washington	38.52409295	-85.9529973
15T-017	OBS-06-0002	South Fork Blue River	Bowers Knob Rd	Washington	38.52047971	-85.93932946
15T-018	OBS-06-0019	Jeff Branch	Bethel Rd	Washington	38.53887871	-85.95606519
15T-019	OBS-06-0011	Honey Run	North Honey Run Rd	Washington	38.52536528	-85.8982047
15T-020	OBS-06-0005	Springle Creek	Blue River Rd	Washington	38.54632127	-85.89874726
15T-021	OBS-06-0010	South Fork Blue River	Casey Hollow Rd	Washington	38.55259127	-85.88819018

<sup>3</sup> 15T-### denotes that these are the selected pour points for this project

## ***I. PROJECT MANAGEMENT/PLANNING***

### ***(QAPP Elements A4, A5, A6, A7, A8)***

#### ***Project/Task Organization and Schedule: (QAPP Element A4)***

The main objective of this project is to provide a comprehensive assessment of the ability of the streams in the South Fork Blue River Watershed to support aquatic life and recreational uses. Sampling for this project will begin in November 2014 and end in October 2015. Barring any hazardous weather conditions or unexpected physical barriers to accessing the site, samples will be collected for physical, chemical, bacteriological parameters, and biological communities.

Timeframes for sampling activities include:

Site reconnaissance activities will be completed in August 2014. Reconnaissance activities will be conducted in the office and through physical site visits.

Water chemistry will be sampled monthly at all sites in the watershed during the recreational season, defined as April through October in the Indiana Administrative Code (IAC, updated October 22, 2014) [327 IAC 2-1-6]. During the months of November through March, only sites at the pour point of each 12 digit HUC will be sampled monthly. The first sampling event will be conducted in November 2014 and the study will conclude in October 2015.

Biological sampling activities will begin in the summer of 2015 and end no later than October 16, 2015. The basin will be sampled for fish community, macroinvertebrate community, and habitat quality at all sites in the watershed. Specific dates for fish community and macroinvertebrate collections cannot be given since sampling may be postponed due to scouring of the stream substrate or in-stream cover caused by a high water event, which would result in non-representative samples.

Bacteriological sampling for *Escherichia coli* (*E. coli*) will take place monthly from April through October of 2015 at all sites in the watershed. In addition, *E. coli* samples will be collected five times from each site at equally spaced intervals over a 30-day period during the recreational season of April to October 2015 to determine a geometric mean.

Stream flow will be quantified over the sampling year at sites designated as “pour points” (Table 1) during the monthly water chemistry sampling in each 12 digit HUC. The first measurement event will be conducted in November 2014 and the study will conclude in October 2015.



## ***Background and Project/Task Description: (QAPP Elements A5, A6)***

The Baseline Monitoring program was instituted to assist in characterizing existing conditions in watersheds throughout the state. The South Fork Blue River baseline data set will be utilized by the TMDL program and shared with local watershed groups and any other interested parties. This monitoring will provide data for TMDL development and watershed planning uses and will aid in the evaluation of future changes within the basin. For this study, the following media will be used for assessment purposes: Water chemistry, stream flow, bacteriological contamination in the form of *E. coli*, fish community, macroinvertebrate assemblages, and habitat evaluations.

## ***Data Quality Objectives (DQOs): (QAPP Element A7)***

The Data Quality Objective (DQO) process (U.S. EPA 2000) is a planning tool for data collection activities. It provides a basis for balancing decision uncertainty with available resources. The DQO is required for all significant data collection efforts for a project. It is a seven-step systematic planning process used to clarify study objectives, define the appropriate types of data, and establish decision criteria on which to base the final use of the data. The DQO for the Baseline Monitoring of the South Fork Blue River is identified in the following seven steps:

### **1. State the Problem**

Indiana is required to assess all waters of the state to determine their designated use attainment status. “Surface waters of the state are designated for full body contact recreation” and “will be capable of supporting” a “well-balanced, warm water aquatic community” [327 IAC 2-1-3]. Data from the intensive sampling of the South Fork Blue River Watershed is needed to develop a TMDL and fully characterize the current water quality condition of the watershed. This project will gather stream flow, water chemistry, bacteriological, biological (fish and macroinvertebrates), and habitat data for the purpose of assessing the designated use attainment status of the South Fork Blue River Watershed.

### **2. Identify the Decision**

The main objective of this study is to fully assess whether the surface waters in this watershed are supporting or non-supporting for aquatic life use and recreational use, and the extent of impairment if they are non-supporting. All sites will be sampled for concentrations of physical, chemical, and biological parameters and evaluated as

“supporting” or “non-supporting” when compared with water quality criteria included below in Table 2 [327 IAC 2-1-6] following Indiana’s 2014 Consolidated Assessment Listing Methodology (CALM, IDEM 2014b).

In addition to the physical, chemical, and bacteriological criteria listed in Table 2, data for several nutrient parameters will be evaluated with the benchmarks described below. Assuming a minimum of three sampling events, if two or more of the conditions below are met on the same date, the waterbody will be classified as non-supporting due to nutrients.

- Total Phosphorus (TP):
  - one or more measurements greater than 0.3 mg/L
- Nitrogen (measured as Nitrate + Nitrite):
  - one or more measurements greater than 10.0 mg/L
- Dissolved Oxygen (DO):
  - any measurement less than 4.0 mg/L;
  - any measurements consistently at or close to the standard, range 4.0-5.0 mg/L; or,
  - any measurement greater than 12.0 mg/L
- pH:
  - any measurement greater than 9.0 Standard Units (SU); or,
  - measurements consistently at or close to the standard, range 8.7-9.0 SU

#### Biological Criteria:

Indiana narrative biological criteria located at 327 IAC 2-1-3 states that “all waters, except as described in subdivision (5),” (i.e. limited use waters) “will be capable of supporting” a “well-balanced, warm water aquatic community.” The water quality standard definition of a “well-balanced aquatic community” is “an aquatic community that: (A) is diverse in species composition; (B) contains several different trophic levels; and (C) is not composed mainly of pollution tolerant species” [327 IAC 2-1-9]. An interpretation or translation of narrative biological criteria into numeric criteria would be as follows: A stream segment is non-supporting for aquatic life use when the monitored fish or macroinvertebrate community receives an Index of Biotic Integrity (IBI) score of less than or equal to 35 which is considered “Poor” or “Very Poor” (IDEM 2014b).

**Table 2. Water Quality Criteria 327 IAC 2-1-6**

Parameters	Water Quality Criteria	Criterion
<i>E. coli</i> April-October (Recreational season)	≤125 MPN/100 mL	5-Sample Geometric Mean
	≤235 MPN/100 mL	Single Sample Maximum
Total Ammonia (NH <sub>3</sub> -N)	Calculated based on pH and Temperature	Calculated CAC
Nitrate+Nitrite-Nitrogen	≤10 mg/L	Human Health point of drinking water intake
Dissolved Oxygen	At least 5.0 mg/L (Warm Waters)	Daily Average
	Not less than 4.0 mg/L at any time	Single Reading
pH	6.0 - 9.0 S.U. except for daily fluctuations that exceed 9.0 due to photosynthetic activity	Single Reading
Temperature	Varies Monthly	1% Annual; Maximum Limits
Chloride	Calculated based on hardness and sulfate	Calculated CAC

MPN = Most Probable Number, CAC = Chronic Aquatic Criterion, S.U. = Standard Units

### 3. Identify the Inputs to the Decision

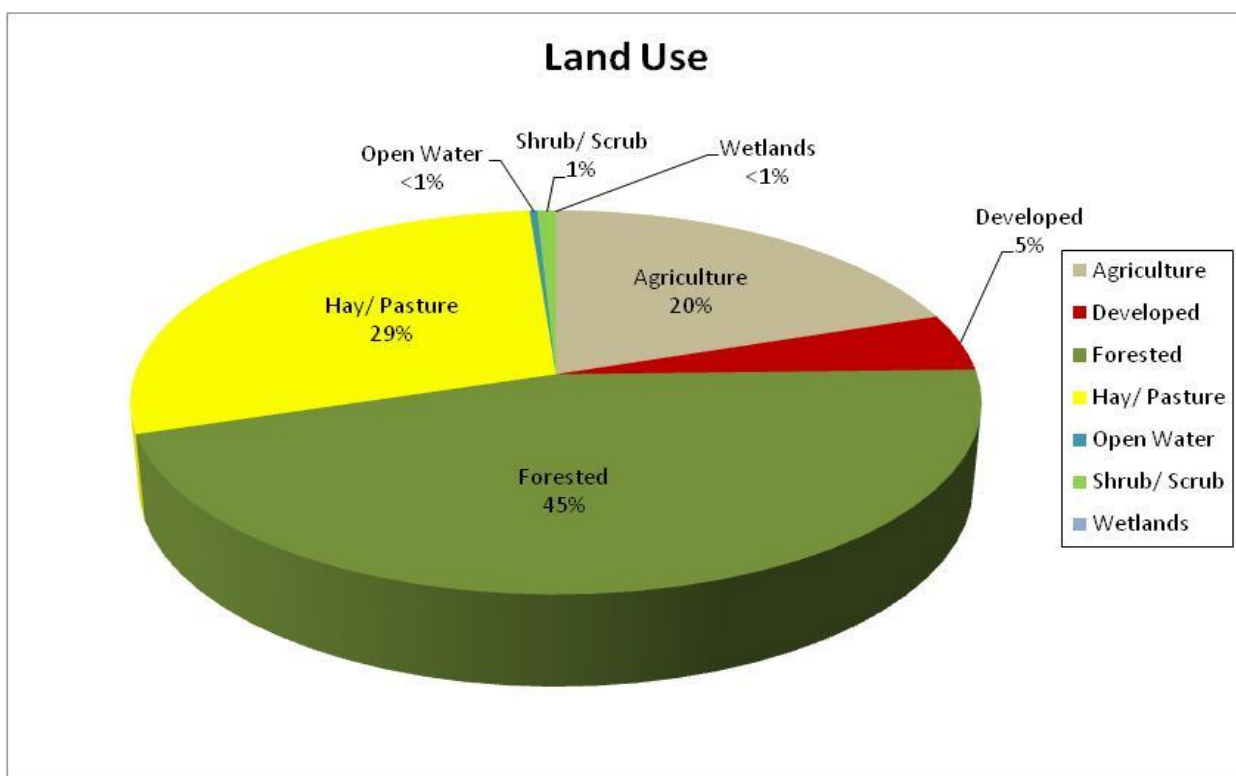
Grab samples will be collected at the surface water sampling locations for *E. coli* and the parameters listed in Table 3. Field measurements (Table 4, page 17) will be conducted at each site during each sampling event. Visual field observations will include weather conditions, stream conditions, and percent stream canopy at each sampling location. All samples collected for bacteriological samples will be analyzed for *E. coli* using the Idexx Colilert Enzyme Substrate Standard Method SM9223B (Clesceri et al., 1998). Surface water chemistry samples will be collected monthly and processed and analyzed by the Indiana State Department of Health (ISDH) Environmental Lab using the analytical methods listed in Table 3. Stream discharge will also be measured monthly at pour points to determine total stream loadings. A fish and macroinvertebrate community sample will be collected once at each site with a corresponding habitat evaluation.

#### 4. Define the Boundaries of the Study

The South Fork Blue River Watershed covers 126 square miles and is located primarily in Clark, Floyd, Harrison, Scott, and Washington counties. The watershed is approximately 45% forested, 29% hay/ pasture, and 20% agriculture. See Figure 2 for the South Fork Blue River Watershed 2012 land use.

See Figure 1 for the South Fork Blue River Watershed Baseline Monitoring sampling area and Table 1 for the list of sampling locations.

**Figure 2. South Fork Blue River Watershed Land Use<sup>2</sup>**



<sup>2</sup>United States Department of Agriculture (USDA) 2012 Crop Data Layer (CDL)

## **5. Develop a Decision Rule**

For assessment purposes in the Indiana Integrated Report (IDEM 2014b), recreational use attainment decisions will be based on bacteriological criteria developed to protect primary contact recreational activities [327 IAC 2-1-6]. Aquatic life use support decisions will include independent evaluations of biological and chemical data as outlined in Indiana's 2014 Consolidated Assessment and Listing Methodology (CALM, IDEM 2014b).

## **6. Specify Tolerable Limits on Decision Errors**

Sampling design error is minimized by utilizing a comprehensive checklist of informational sources, evaluation of historical information, and a thorough watershed pre-survey. This sampling design has been formulated to address data deficiencies and render the optimum amount of data needed to fill gaps in the decision process.

Good quality data are essential for minimizing decision error. By minimizing both sampling design error and measurement error for physical and biological parameters, more confidence can be placed in the conclusions drawn on the stressors and sources affecting the water quality in the study area.

Site specific aquatic life use and recreational use assessments include program specific controls to minimize the introduction of errors. These controls include: water chemistry and bacteriological blanks and duplicates, biological site revisits or duplicates, and laboratory controls through verification of species identifications. Field Procedure Manuals (IDEM 2002; OHEPA 2006) and Standard Operating Procedures (IDEM 1992b, 1992c, 1992d, 1992e, 2010a) dictate consistent and proven techniques for sample collection to assure representative samples and minimize measurement error.

The QA/QC process detects deficiencies in the data collection as set forth in the IDEM QAPP for the Indiana Surface Water Quality Monitoring Program (IDEM 2004). The QAPP requires all contract laboratories to adhere to rigorous standards during sample analyses and to provide good quality usable data. Chemists within the Watershed Assessment and Planning Branch (WAPB) review the laboratory analytical results for quality assurance. Any data which is "Rejected" due to analytical problems or errors will not be used for water quality assessment decisions. Any data flagged as "Estimated" may be used on a case-by-case basis.

## **7. Optimize the Design for Obtaining Data**

A Modified Geometric Design (OHEPA 1999, 2012) site selection process (Attachment 1) is used in this study to get the necessary spatial representation of the entire study area. Sites within this watershed have been selected based on a geometric progression

of drainage areas and then located to the nearest bridge. Sample sites at road crossings allows for more efficient sampling of the watershed.

### ***Training and Staffing Requirements: (QAPP Element A8)***

The WAPB uses many Standard Operating Procedures (SOPs), so any new staff member must be trained by experienced IDEM professionals on how to operate field and laboratory equipment for the collection of chemical, physical, and biological parameters as well as how to perform required QA/QC procedures (information about SOPs is given in Sections II MEASUREMENT/DATA ACQUISITION and IV DATA VALIDATION and USABILITY). Before sampling starts, IDEM staff spend several days reviewing SOPs with field and laboratory personnel that may be involved with the project.

The fish or macroinvertebrate community field Crew Chief must have a Bachelor of Science degree with a concentration in biology or other closely related area and at least one year of experience with the sampling methodology and taxonomy of the aquatic communities in the region. Prior to conducting electrofishing for fish community sampling, all crew members should review the Principles and Techniques of Electrofishing correspondence course provided by the U.S. Fish & Wildlife Service, National Conservation Training Center. Field Crew Chiefs will test electrofishing equipment and conduct field training with less experienced crew members. The field Crew Chief will be responsible for completion of field data sheets, taxonomic accuracy, sampling efficiency and representation, and voucher specimen tracking.

Staff from the Technical and Logistical Services Section will assist with laboratory work requests and review laboratory data for adherence to QA/QC requirements specified in analytical test methods, contract requirements, and the IDEM QAPP for the Indiana Surface Water Quality Monitoring Program (IDEM 2004) as well as importing electronic data into the Assessment Information Management System (AIMSII) database which is used by the WAPB. The Quality Assurance Officer will create QA/QC review reports for each laboratory analysis set. Quality Assurance staff will conduct audits of field sampling procedures utilized by WAPB staff. Monitoring staff will oversee the entry of the field and laboratory data into AIMSII and perform data QA/QC for accuracy and completeness.

## ***II. Measurement/Data Acquisition***

### ***Sampling Process Design/ Methods, Sample Handling and Custody (QAPP Elements B1, B2, B3, B4, B5, B6, B7)***

#### **Sampling Sites/Sampling Design: (QAPP Element B1)**

The proposed site locations are chosen using a modified geometric and targeted design as described previously in the “Baseline Monitoring Program Objective” section of this workplan.

Site reconnaissance activities are conducted in-house and through physical site visits. In-house activities include preparation and review of site maps and aerial photographs. Physical site visits include verification of accessibility, safety considerations, equipment needed to properly sample the site, and property owner consultations, if required. All information will be recorded on the IDEM Site Reconnaissance Form (Attachment 2) and entered into the AIMS II database. Final coordinates for each site will be determined during the physical site visits or at the beginning of the sampling phase of this project using a Trimble Juno <sup>TM</sup> SB Global Positioning System (GPS), with an accuracy of one to three meters. These coordinates will be entered into the AIMS II database.

Table 1 provides a list of the selected sampling sites with the stream name, AIMS Site Number, County Name, and the latitude and longitude of each site. The map at Figure 1, paired with that table, provides a good overview of the various sampling site locations.

#### **Sampling Methods and Sample Handling: (QAPP Elements B2, B3)**

##### **Water Chemistry**

One team of two staff will collect grab water chemistry samples and record physical site observations on the stream sampling field data sheet (Attachment 3), during monthly sampling events. All water chemistry sampling will adhere to the Water Quality Surveys Section Field Procedure Manual Section 2.1 (IDEM 2002).

**Bacteriological Sampling** The bacteriological sampling will be conducted by one team consisting of one or two staff. Samples will be processed in an IDEM Fixed and/or Mobile *E. coli* Laboratory equipped with all materials and equipment necessary for the Colilert® Test Method. Per Element A4 Project Organization and Schedule (above), the expected time frame for bacteriological sampling will be April through October of 2015. Staff will collect the samples in a 120 mL pre-sterilized wide-mouth container from the center of flow if stream is wadeable or from the shoreline using a pole sampler if the stream is not wadeable. All samples will be consistently labeled, cooled, and held at a temperature less than 10°C during transport. All *E. coli* samples will be collected on a schedule such that any sampling crew can deliver them to the appropriate IDEM *E. coli* Laboratory for analyses within the bacteriological holding time of six hours.

The IDEM Mobile *E. coli* Laboratory is used in this project to facilitate *E. coli* testing by eliminating the necessity of transporting samples to distant contract laboratories within a six hour holding time. The IDEM Mobile *E. coli* Laboratory (Van) provides work space containing storage for samples, supplies for Colilert® Quanti-tray testing, and all equipment needed for collecting, preparing, incubating, and analyzing results in the same manner as the IDEM Fixed *E. coli* Laboratory. All supplies will be obtained from IDEXX Laboratories, Inc., Westbrook, Maine.

### **Fish Community Sampling**

The fish community sampling will be completed by teams of three to five staff. Sampling will be performed using various standardized electrofishing methodologies depending on stream size and site accessibility. Fish assemblage assessments will be performed in a sampling reach of 15 times the length of the average wetted width, with a minimum reach of 50 meters and a maximum reach of 500 meters (Simon 1997; Simon and Dufour 1998; U.S. EPA 1995). An attempt will be made to sample all habitat types available within the sample reach to ensure adequate representation of the fish community present at the time of the sampling event. The possible list of electrofishers to be utilized include: the Smith-Root LR-24 or LR-20 Series backpack electrofishers; the Smith-Root model 1.5KVA electrofishing system; the Smith-Root model 2.5 Generator Powered Pulsator electrofisher with RCB-6B junction box and rat-tail cathode cable assembled in a canoe (if parts of the stream are not wadeable, the system may require the use of a dropper boom array outfitted in a canoe or possibly a 12 foot Loweline™ boat); or, for non-wadeable sites, the Smith-Root model 6a electrofisher assembled in a 16 foot Loweline™ boat (IDEM 1992a, 1992b, 1992c, 1992d).

Sample collections during high flow or turbid conditions will be avoided due to 1) low collection rates which result in non-representative samples and 2) safety considerations



for the sampling team. Sample collections during late autumn and seasonal cold temperatures will be avoided due to the lack of responsiveness to the electrical field by some species that can also result in samples that are not representative of the streams fish assemblage (Simon 1990; U.S. EPA 1995).

Fish will be collected using dip nets with fiberglass handles and netting of 1/8-inch bag mesh. Fish collected in the sampling reach will be sorted by species into baskets and buckets. Young-of-the year fish, less than 20 millimeters (mm) total length, will not be retained in the community sample (Simon 1990; U.S. EPA 1995).

Prior to processing fish specimens and completion of the fish collection datasheet (Attachment 4), one to two individuals per species will be preserved in 3.7% formaldehyde solution for future reference if there are more than 10 individuals for that species collected in the sampling reach, the specimens can be positively identified, and the individuals for preservation are small enough to fit in a 2000 mL jar. If however, there are few individuals captured or the specimens are too large to preserve, a photo of key characteristics will be taken for later examination. Taxonomic characteristics for possible species encountered in the basin of interest will be reviewed prior to field work. Fish specimens should also be preserved if they cannot be positively identified in the field (especially those that co-occur like the striped and common shiner), individuals that appear to be hybrids or have anomalies, as well as dead specimens that are taxonomically valuable for un-described taxa (like the new stoneroller, red shiner, or jade darter), life history studies, or research projects.

Data will be recorded for non-preserved fish on the fish collection datasheet (Attachment 4) consisting of the following: number of individuals, minimum and maximum total length in millimeters (mm), mass weight in grams (g), and number of individuals with deformities, eroded fins, lesions, tumors, and other anomalies. Once the data have been recorded, specimens will be released within the sampling reach if possible. Data will be recorded for preserved fish specimens following taxonomic identification in the laboratory.

### **Macroinvertebrate Sampling**

The macroinvertebrate community sampling may be conducted immediately following the fish community sampling event or on a different date by crews of two to three staff. Samples are collected using a modification of the U.S. Environmental Protection Agency (EPA) Rapid Bioassessment Protocol multi-habitat (MHAB) approach using a D-frame dipnet (Barbour et al. 1999; IDEM 2010a; Klemm et al. 1990; Plafkin et al. 1989). The IDEM MHAB approach (IDEM 2010a) is composed of a 1-minute "kick" sample within a riffle or run (collected by disturbing one square meter of stream bottom substrate in a riffle or run habitat and collecting the dislodged macroinvertebrates within

the dipnet) and a 50 meter “sweep” sample of shoreline habitats (collected by disturbing habitats such as emergent vegetation, coarse particulate organic matter, depositional zones, logs and sticks and collecting the dislodged macroinvertebrates within the dipnet). The 50 meter length of riparian corridor that is sampled at each site will be defined using a rangefinder or GPS unit. If the stream is too deep to wade, a boat will be used to sample the 50 meter zone along the shoreline that has the best available habitat. The 1-minute “kick” and 50 meter “sweep” samples are combined in a bucket of water which will be elutriated through a U.S. standard number 35 (500 µm) sieve a minimum of five times so that all rocks, gravel, sand and large pieces of organic debris are removed from the sample. The remaining sample is then transferred from the sieve to a white plastic tray where the collector (while still on-site) will conduct a 15-minute pick of macroinvertebrates at a single organism rate with an effort to pick for maximum organism diversity through turning and examination of the entire sample in the tray. The resulting picked sample will be preserved in 70% isopropyl alcohol and returned to the laboratory for identification at the lowest practical taxonomic level (usually genus or species level, if possible) and evaluated using the MHAB macroinvertebrate IBI. Before leaving the site, an IDEM OWQ Macroinvertebrate Header Form (Attachment 5) will also be completed for the sample. A completed Biological Samples’ chain-of- custody form (Attachment 6) accompanies the samples through the identification process.

### **Habitat Assessments**

Habitat assessments will be completed immediately following macroinvertebrate and fish community sample collections at each site using a slightly modified version of the Ohio Environmental Protection Agency (OHEPA) Qualitative Habitat Evaluation Index (QHEI), 2006 edition (OHEPA 2006; Rankin 1995). A separate QHEI (Attachment 7) must be completed for these two media since the sampling reach length is different (i.e., 50 meters for macroinvertebrates and between 50 and 500 meters for fish).

### **Field Parameter Measurements**

Dissolved oxygen (DO), pH, water temperature, specific conductance, and DO percent saturation will be measured with a datasonde during each sampling event, regardless of the media type being collected (IDEM 2002). Measurement procedures and operation of the datasonde shall be performed according to the manufacturers’ manuals (Hydrolab Corporation 2002; YSI 2002) and Sections 2.10 – 2.13 of the Water Quality Surveys Section Field Procedure Manual (IDEM 2002). Turbidity will be measured with a Hach™ turbidity kit, and the meter number written in the comments under the field parameter measurements. If a Hach™ turbidity kit is not available, the datasonde measurement for turbidity will be recorded. All field parameter measurements will be recorded on the IDEM Stream Sampling Field Data Sheet (Attachment 3).

## **Flow Measurements**

Flow measurements are to be taken by the water chemistry crew at the pour point sites during each sampling run using the SonTek Acoustic Doppler Profiler (ADP) at non-wadeable sites and the FlowTracker Handheld Acoustic Doppler Velocimeter (ADV)®, Ott Acoustic Digital Current (ADC), or Ott MF pro at the wadeable sites. Procedures shall be according to Section 2.6.5 of the Surveys Section Field Procedure Manual (IDEM 2002) and the manufacturers' operating manuals. (SonTek/YSI Inc 2007; 2001)

## ***Analytical Methods: (QAPP Element B4)***

### **Laboratory Procedure for *E. coli* Measurements:**

At the end of each sampling run and while still in the field, water samples are processed and analyzed for *E. coli* within the six-hour holding time for collection and transportation, and the two-hour holding time for sample processing. All waters sampled are processed and analyzed for *E. coli* in the IDEM *E. coli* Mobile Laboratory or IDEM Shadeland laboratory, which is equipped with required materials and equipment necessary for the Idexx<sup>TM</sup> Colilert Test. The Colilert Test is a multiple-tube Enzyme Substrate Standard Method SM-9223 B (Clesceri et al., 1998). The *E. coli* test method and quantification limit are identified below in Table 3.

### **Nutrient and General Chemistry Parameters Measurements:**

Nutrient and general chemistry measurement analysis is performed at ISDH Environmental Lab in accordance with pre-approved test methods and allotted time frames. The nutrient and general chemistry parameters and their respective test methods and quantification limits are identified below in Table 3. A chain-of-custody form created by the AIMS II database (Attachment 8) and a sample analysis request form (Attachment 9) accompanies each sample set through the analytical process.

**Table 3. *E. coli*, Nutrient and General Chemistry Parameters Test Methods**

Parameter	Method	Limits of Quantification	Units	Preservative	Holding Times
<i>E. coli</i>	SM-9223 B Enzyme Substrate Test	1.0	*MPN /100 mL	0.0008% Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub> for CL <sub>2</sub>	8 hours
Alkalinity (as CaCO <sub>3</sub> )	SM 2320B	10.0	mg/L	None	14 days
Total Solids	SM 2540B	10.0	mg/L	None	7 days
Total Suspended Solids	SM 2540D	4.0	mg/L	None	7 days
Total Dissolved Solids	SM 2540C	10.0	mg/L	None	7 days
Sulfate	EPA 300.0	.3	mg/L	None	28 days
Chloride	EPA 300.0	.25	mg/L	None	28 days
Hardness (as CaCO <sub>3</sub> )	SM 2340B	1.0	mg/L	HNO <sub>3</sub> < pH 2	6 months
Ammonia Nitrogen	SM 4500NH <sub>3</sub> -D	0.10	mg/L	H <sub>2</sub> SO <sub>4</sub> < pH 2	28 days
TKN	ASTM D3590-89	0.30	mg/L	H <sub>2</sub> SO <sub>4</sub> < pH 2	28 days
Nitrate/Nitrite	EPA 353.2	0.05	mg/L	H <sub>2</sub> SO <sub>4</sub> < pH 2	28 days
Total Phosphorus	SM 4500P-E	0.05	mg/L	H <sub>2</sub> SO <sub>4</sub> < pH 2	28 days
TOC	SM 5310C	1.0	mg/L	H <sub>2</sub> SO <sub>4</sub> < pH 2	28 days
COD	EPA 410.4	10.0	mg/L	H <sub>2</sub> SO <sub>4</sub> < pH 2	28 days

\* Clesceri et al., 1998. 1 MPN = 1 CFU/100 mL

### Field Parameters Measurements:

The field measurements of DO, temperature, pH, conductivity, and turbidity are taken each time a sample is collected. The field parameters and their respective test methods and sensitivity limits are identified below in Table 4.

During each sampling run, field observations from each site and ambient weather conditions at the time of sampling are noted and documented on stream sampling field data sheets (Attachment 3). Digital photos up-stream and down-stream of the sampling site will be taken, logged, and documented for later references.

**Table 4. Field Parameters Test Methods**

Parameter	Method	Sensitivity Limit	Units
Dissolved Oxygen (Datasonde optical)	ASTM D888-09(C)	0.01	mg/L
Dissolved Oxygen (Winkler Titration)	SM 4500-OC <sup>1</sup>	0.2	mg/L
Dissolved Oxygen % Saturation (Datasonde optical)	ASTM D888-09(C)	0.01	%
Turbidity (Datasonde)	SM2130B	0.02	NTU
Turbidity (Hach Turbidimeter)	EPA 180.1 <sup>1</sup>	0.01	NTU
Specific Conductance (Datasonde)	SM 2510B	1.0	μS/cm
Temperature (Datasonde)	SM 2550B(2)	0.1	° C
Temperature (field meter)	SM 2550B(2) <sup>1</sup>	0.1	° C
pH (Datasonde)	EPA 150.2	0.01	SU
pH (field meter)	SM 4500-HB <sup>1</sup>	0.01	SU

<sup>1</sup> Method used for Field Calibration Verification

### ***Quality Control and Custody Requirements: (QAPP Element B5)***

Quality assurance protocols will follow part B5 of the “Quality Assurance Project Plan for the Indiana Surface Water Quality Monitoring and Total Maximum Daily Load (TMDL) Program,” Revision 3, by Timothy Bowren and Dr. Syed Ghiasuddin (IDEM 2004).

## ***Field Instrument Testing and Calibrations: (QAPP Elements B6, B7)***

The Datasonde will be calibrated immediately prior to each week's sampling (IDEM 2002). Calibration results and drift values will be recorded, maintained, stored, and archived in log books located in the calibration laboratories at the Shadeland facility. The drift value is the difference between two successive calibrations. Field parameter calibrations will conform to the procedures as described in the instrument users' manuals (Hydrolab Corporation 2002; YSI 2002). The DO component of the calibration procedure will be conducted using the air calibration method. The unit will be field checked for accuracy once during the week by comparison with a Winkler DO test, as well as Hach™ turbidity, pH, and temperature meters. Weekly calibration verification results will be recorded on the stream sampling field data sheets (Attachment 3) and entered into the AIMS II database. A Winkler DO test will also be conducted at sites where the DO concentration is 4.0 mg/L or less.

### **Field Analysis Data**

*In-situ* water chemistry field data are collected in the field using calibrated or standardized equipment. Calculations may be done in the field or later at the office. Analytical results, which have limited QC checks, are included in this category. Detection limits and ranges have been set for each analysis (Table 4). Quality control checks (such as duplicate measurements, measurements of a secondary standard, or measurements using a different test method or instrument) which are performed on field or laboratory data are usable for estimating precision, accuracy, and completeness for the project.

### **Bacteriological Sampling**

Bacteriological samples will be analyzed using the SM 9223 Enzyme Substrate Coliform Test Method, see Table 3 for quantification limits. Samples will be collected using 120 mL pre-sterilized wide-mouth containers and adhere to the six-hour holding time. Analytical results from an IDEM Fixed and/or Mobile *E. coli* Laboratory include QC check sample results from which precision, accuracy, and completeness can be determined for each batch of samples. Raw data are archived by analytical batch for easy retrieval and review. Chain-of-custody procedures must be followed, including: time of collection, time of setup, time of reading the results, and time and method of disposal (IDEM, 2002). Any method deviations will be thoroughly documented in the raw data. All QA/QC samples will be tested according to the following guidelines:

Field Duplicate:      Field Duplicates will be collected at a frequency of one per batch or at least one for every 20 samples collected ( $\geq 5\%$ ).

- Field Blank: Field Blanks will be collected at a frequency of one per batch or at least one for every 20 samples collected ( $\geq 5\%$ ).
- Laboratory Blank: Laboratory Blanks (sterile laboratory water blanks) will be tested at a frequency of one per day.
- Positive Control: Each lot of media will be tested for performance using *E. coli* bacterial cultures.
- Negative Controls: Each lot of media will be tested for performance using non-*E. coli* and noncoliform bacterial cultures.

### **Water Chemistry Data**

Sample bottles and preservatives used will be certified for purity by the manufacturer. Sample collection containers for each parameter, preservative and holding time (Table 3) will adhere to U.S. EPA requirements. Field duplicates and matrix spike/matrix spike duplicates (MS/MSD) shall be collected at the rate of one per sample analysis set or one per every 20 samples, whichever is greater. Additionally, field blank samples will be taken at a rate of one set per sample analysis set or one per every 20 samples, whichever is greater.

### **Fish Community Data**

Replicate fish community sampling will be performed at a rate of 10 percent of the total fish community sites sampled, or approximately four in the basin (U.S. EPA 1995). Replicate sampling will be performed once all initial sites have been sampled, with at least two weeks of recovery between the initial and replicate sampling events. The fish community replicate sampling and habitat assessment will be performed with either a partial or complete change in field team members (U.S. EPA 1994; U.S. EPA 1995). The resulting IBI and QHEI total score between the initial visit and the revisit will be used to evaluate precision. A chain-of-custody form is used to track samples from the field to the laboratory (Attachment 6). Fish in the laboratory may be verified by regionally recognized non-IDEM freshwater fish taxonomists. All data are checked for:

- 1) completeness
- 2) calculations performed
- 3) data entered into the AIMS II database
- 4) checked again for data entry errors.

### **Macroinvertebrate Community Data**

Replicate macroinvertebrate field samples will be collected at every 10<sup>th</sup> site. This will result in a precision evaluation based on a 10% replicate of samples collected. Records

of laboratory identifications and the QA/QC of taxonomic work is maintained by the laboratory supervisor of the Probabilistic Monitoring Section of IDEM.

### ***III. ASSESSMENT/OVERSIGHT: (QAPP Elements C1, C2)***

Field and laboratory performance and system audits will be performed to ensure good quality data. The field and laboratory performance includes precision measurements by relative percent difference of field and laboratory duplicate, accuracy measurements by percent of recovery of MS/MSD samples analyzed in the laboratory, and completeness measurements by the percent of planned samples that are actually collected, analyzed, reported, and usable for the project.

#### ***Data Quality Assessment Levels***

The samples and various types of data collected by this program are intended to meet different DQA Levels as cited in the QAPP for Indiana Surface Water Quality Monitoring Program, Revision 3 (IDEM 2004). The level of Quality Assurance and the DQA Level to which the analytical data qualifies will be as follows:

**DQA Level 1 Screening Data:** The results are usually generated onsite and have no QC checks. Analytical results, which are just numbers, and have no QC checks, no precision or accuracy information, and no detection limit calculations are included in this category. Onsite data are primarily used for pre-surveys and for preliminary rapid assessment.

**DQA Level 2 Field Analysis Data:** Data are recorded in the field or laboratory on calibrated or standardized equipment. Field duplicates are measured on a regular periodic basis. Calculations may be done in the field or later at the office. Analytical results, which have limited QC checks, are included in this category. Detection limits and ranges have been set for each analysis. The QC checks information for field or laboratory results is useable for estimating precision, accuracy, and completeness for the project. Data from this category are used independently for rapid assessment and preliminary decisions.

**DQA Level 3 Laboratory Analytical Data:** Analytical results include QC check samples for each batch of samples from which precision, accuracy, and completeness can be determined. Method detection limits (MDLs) have been determined using 40 Code of Federal Regulations (CFR) Part 136 Appendix B (CFR 2013). Additionally, all reporting information required in the laboratory contract and in the IDEM Surface Water Quality Monitoring and TMDL QAPP, especially Table



A9-1, are included in the analytical data reports. Raw data, chromatograms, spectrograms, and bench sheets are not included as part of the analytical report, but are maintained by the contract laboratory for easy retrieval and review. Data can be elevated from DQA Level 3 to DQA Level 4 by inclusion of this information in the data report and the QC data are reported using contract laboratory program (CLP) forms or CLP format. Data in this category are considered as complete, legally defensible, and used for regulatory decisions.

**DQA Level 4 Enforcement Data:** *Analytical results mostly meet the U.S. EPA required CLP data analysis, Contract Required Quantification Limits (CRQL), and validation procedures.* QC data are reported on CLP forms or CLP format. Raw data, chromatograms, spectrograms, and bench sheets are included as part of the analytical report. Additionally, all reporting information required in the laboratory contract, and in the *IDEM Surface Water Quality Monitoring Program and TMDL QAPP*, are included in the analytical data reports. Data falling under this category are considered as complete, legally quantitative in value, and used for regulatory decisions.

All samples collected for bacteriological and laboratory analysis for this project will adhere to DQA Level 3. All field parameters collected for this project will adhere to DQA Level 2. All of the sample data are QA/QC'd for completeness, precision, and accuracy.

#### **IV. DATA VALIDATION AND USABILITY: (QAPP Element D1, D2)**

##### **Quality Assurance/Data Qualifiers and Flags:**

The various data qualifiers and flags used for Quality Assurance and validation of the data are outlined below in Table 5.

**Table 5. Data Qualifiers and Flags**

<b>Flags</b>	<b>Description</b>
<b>R</b>	<b>Rejected.</b> Result is not acceptable for use in decision making processes.
<b>J</b>	<b>Estimated.</b> The use of the result in decision-making processes will be determined on a case-by-case basis.

Flags	Description
<b>U</b>	<b>Between MDL and RL</b> -- The result of the parameter is above the Method Detection Limit (MDL) but below the Lab Reporting Limit (RL) and will be estimated.
<b>Q</b>	<b>QC Checks or Criteria</b> -- One or more of the QC checks or criteria is out-of-control.
<b>D</b>	<b>RPD for Duplicates</b> -- The Relative Percent Difference (RPD) for a parameter is outside the acceptable control limits. The parameter will be considered estimated or rejected on the basis listed below: <ol style="list-style-type: none"> <li>1. If the Sample or Duplicate value is less than the RL, and the other value exceeds 5 times the MDL, then the sample will be estimated.</li> <li>2. If the RPD is outside the established control limits (max. RPD) but below two times the established control limits (max. RPD), then the sample will be estimated.</li> <li>3. If the RPD is twice the established control limits (max. RPD) or greater, then the sample will be rejected.</li> </ol>
<b>B</b>	<b>Blank Contamination</b> -- This parameter is found in a field or a lab blank. Whether the result is accepted, estimated, or rejected will be based upon the level of contamination listed below: <ol style="list-style-type: none"> <li>1. If the result of the sample is greater than the reporting limit but less than five times the blank contamination, the result will be rejected.</li> <li>2. If the result of the sample is between five and ten times the blank contamination, the result will be estimated.</li> <li>3. If the result of the sample is less than the reporting limit or greater than ten times the blank contamination, the result will be accepted.</li> </ol>
<b>H</b>	<b>Holding Time</b> -- The analysis for this parameter was performed out of the holding time. The results will be estimated or rejected on the basis listed below: <ol style="list-style-type: none"> <li>1. If the analysis was performed between the holding time limit and 1.5 times the holding time limit, the result will be estimated.</li> <li>2. If the analysis was performed outside the 1.5 times the holding time limit, the result will be rejected.</li> </ol>

### ***Data Usability:***

The environmental data collected and its usability are finally qualified and classified into one or more of the four categories: Acceptable Data, Enforcement Capable Results, Estimated Data, and Rejected Data.

- **Acceptable Data** are suitable for decision making and have no flagged data points.
- **Enforcement Capable Results** meets all QC checks and have no flagged data points.
- **Estimated Data** may be suitable for enforcement or decision making on a case by case basis.
- **Rejected Data** are not suitable for enforcement or for decision making.

### ***Laboratory and Estimated Cost:***

Laboratory analysis and data reporting for this project will comply with the QAPP for Indiana Surface Water Quality Monitoring and TMDL Program (IDEM/100/29/338/073/2004, see IDEM 2004), Request for Proposals (RFP) 12-48 (see IDEM 2012), and the Office of Water Quality Assessment Branch Quality Management Plan, see IDEM 2008a). Analytical tests on the general chemistry and nutrient parameters outlined in Table 3 will be performed by the ISDH Environmental Lab in Indianapolis, Indiana at no direct cost. Supplies for the bacteriological sampling will come from IDEXX Laboratories, Inc., Westbrook, Maine with a total estimated cost for this project of \$1,280. All fish and macroinvertebrate samples will be collected and analyzed by IDEM staff.

### ***Reference Manuals and Personnel Safety:***

All staff persons who participate in the field component of this study are required to have completed Basic First Aid and Cardio-Pulmonary Resuscitation (CPR) training. According to the memorandum "Change in status of Water Assessment Branch staff in accordance with the Agency training policy," dated November 29, 2010, OWQ Watershed Assessment and Planning Branch staff is exempt from initial and annual training requirements set forth in Section 6.0 of the IDEM Health and Safety Training Policy (IDEM 2010b). The memorandum also states "as an alternative to the training requirements of the policy, the WAPB will conduct in-service training at a minimum of four (4) hours per year on topics directly related to duties performed by staff." New hires or those changing job responsibilities without the minimum four-hour training must be accompanied in the field by a staff member who has met the requirements of the branch Health and Safety training.

Field personnel collecting water chemistry and bacteriological samples will follow policies and procedures established in the Surveys Section Field Procedures Manual (IDEM 2002) and the Hazardous Communication Plan Supplement (IDEM 1997). Field

personnel collecting fish and macroinvertebrate community samples must read and comply with the Biological Studies Section SOP Manual: Section II. Hazard Communications Manual (IDEM 1992e) which includes four yellow three-ring binders consisting of the:

- 1) WAPB Safety Manual;
- 2) IDEM Hazard Communications SOP;
- 3) Occupational Safety and Health Administration Handbooks;
- 4) Material Safety Data Sheets;
- 5) "Field and Laboratory Operating Procedures for use, handling and storage of chemicals in the laboratory" (Newhouse 1998a); and,
- 6) "Field and Laboratory Operating Procedures for Use, Handling, and Storage of Solutions Containing Formaldehyde" (Newhouse 1998b).

Sampling on surface waters requires safety consciousness of staff members and the use of specialized equipment; thus, staff will comply with the IDEM Personal Protective Equipment (PPE) Policy (IDEM 2008b). If an injury or illness arises in the field, staff will follow the IDEM Injury and Illness Resulting from Occupational Exposure Policy (IDEM 2010c).

Operating in and around waterbodies carries inherent risks of drowning; thus, personnel involved in sample collection will wear appropriate clothing and PPE when operating boats or sampling in deep water or swift currents. According to the memorandum "Use of Personal Flotation Devices (PFDs) by Branch Personnel," dated February 29, 2000, WAPB staff must wear U.S. Coast Guard approved Type I, II, or III PFDs whenever:

- the planned work requires them to enter the water and the maximum water depth at any portion of the work site is over their knee (note that this depth depends on the employee but it will usually be between 12 and 20 inches or 300-500 mm);
- the employee is in a watercraft of any kind that is being launched, is in the water, or is being retrieved from the water; or,
- the employee must work from structures that do not possess guard rails and are over or alongside water where the water depth is or could reasonably be expected to be three feet deep or greater.

In addition, when work is being done in boats on co-jurisdictional waters (as defined by Indiana Code (IC) 14-8-2-315) or during hours of darkness on any waters of the state, all personnel in the watercraft must wear a high intensity whistle and Safety of Life at Sea (SOLAS) certified strobe light.

Safety issues are the responsibility of all crew members; however, any questions in the field should be directed to the field crew leader. The field crew leader is responsible for the completion of all work listed in the workplan, the health and safety aspects of the sampling event, and successful interactions with landowners and members of the public.



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- IDEM. 1992b, revision 1. Section 4, Standard Operating Procedures for Fish Collections, Use of Seines, Electrofishers, and Sample Processing. Biological Studies Section, Surveillance and Standards Branch, Office of Water Management, Indiana Department of Environmental Management, Indianapolis, Indiana.
- IDEM. 1992c, revision 1. Section 5, Standard Operating Procedures for Conducting Rapid Assessment of Ambient Water Quality Using Fish (RBP-V). Biological Studies Section, Surveillance and Standards Branch, Office of Water Management, Indiana Department of Environmental Management, Indianapolis, Indiana.
- IDEM. 1992d, revision 1. Section 11, Standard Operating Procedures-Appendices of Operational Equipment Manuals and Procedures. Biological Studies Section, Surveillance and Standards Branch, Office of Water Management, Indiana Department of Environmental Management, Indianapolis, Indiana.
- IDEM. 1992e, revision 1. Section 2, Biological Studies Section Hazards Communications Manual (List of Contents). Biological Studies Section, Surveillance and Standards Branch, Office of Water Management, Indiana Department of Environmental Management, Indianapolis, Indiana. (This Manual is not available in

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[&MaximumDocuments=1&FuzzyDegree=0&ImageQuality=r75g8/r75g8/x150y150g16/i425&Display=p%7Cf&DefSeekPage=x&SearchBack=ZyActionL&Back=ZyActionS&BackDesc=Results%20page&MaximumPages=1&ZyEntry=1&SeekPage=x&ZyPURL](#)

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## **Attachment 1: Modified Geometric Design Steps for Baseline Studies**

### **Introduction**

A relatively new design that has recently been implemented in Indiana is termed the Geometric Site Selection process. This design is employed within watersheds that correspond to the 12-14 digit HUC scale in order to fulfill multiple water quality management objectives, not just the conventional focus on status assessment. It is employed at a spatial scale that is representative of the scale at which watershed management is generally being conducted.

Sites within the watershed are allocated based on a geometric progression of drainage areas starting with the area at the mouth of the main stem river or stream (pour point) and working “upwards” through the various tributaries to the primary headwaters. This approach allocates sampling sites in a semi-random fashion and according to the stratification of available stream and river sizes based on drainage area. The Geometric Site Selection process is then modified by adding a targeted selection of additional sampling sites that are used to focus on localized management issues such as point source discharges, habitat modifications, and other potential impacts within a watershed. These sites are then “snapped to bridges” to facilitate safe and easy access to the stream. This design also fosters data analysis that takes into consideration overlying natural and human caused influences within the streams of a watershed. The design has been particularly useful for watersheds that are targeted for TMDL development because missing, incomplete, or outdated assessments can be addressed prior to TMDL development.

## Selection Process

In ArcGIS, download from NHD Plus site (<http://www.horizon-systems.com/nhdplus/HSC-wthMS.php>) the following files for Region 5 (and then again for Region 7) and zip them into the appropriate file structure.

File Description	File Name (.zip***)	Format
Region 05, Version 01_01, Catchment Grid	NHDPlus05V01_01_Catgrid	ESRI Grid
Region 05, Version 01_01, Catchment Shapefile	NHDPlus05V01_01_Catshape	Shapefile
Region 05, Version 01_02, Catchment Flowline Attributes	NHDPlus05V01_02_Cat_Flowline_Attr	DBF
Region 05, Version 01_02, Elevation Unit a	NHDPlus05V01_02_Elev_Unit_a	ESRI Grid
Region 05, Version 01_02, Elevation Unit b	NHDPlus05V01_02_Elev_Unit_b	ESRI Grid
Region 05, Version 01_02, Elevation Unit c	NHDPlus05V01_02_Elev_Unit_c	ESRI Grid
Region 05, Version 01_01, Flow Accumulation and Flow Direction Unit a	NHDPlus05V01_01_FAC_FDR_Unit_a	ESRI Grid
Region 05, Version 01_01, Flow Accumulation and Flow Direction Unit b	NHDPlus05V01_01_FAC_FDR_Unit_b	ESRI Grid
Region 05, Version 01_01, Flow Accumulation and Flow Direction Unit c	NHDPlus05V01_01_FAC_FDR_Unit_c	ESRI Grid
Region 05, Version 01_02, National Hydrography Dataset	NHDPlus05V01_03_NHD	Shapefile and DBF
Region 05, Version 01_01, Stream Gage Events	NHDPlus05V01_01_StreamGageEvent	Shapefile
Region 05, Version 01_01, QAQC Sinks Spreadsheet	NHDPlus05V01_01_QAQC_Sinks	Excel Spreadsheet

Create a new point shapefile (or geodatabase featureclass) named Geometric Design within ArcCatalog with the same projection as the unzipped layers above.

Within an ArcMap project, add the following:

- nhdflopline layer;
- Geometric Design layer;
- catchment shapefile;
- the FlowlineAttributesFlow table.

Add the following fields to the nhdflopline layer:

- LENGTHMi (type: double, precision: 9, scale 4)
- DrainMi (type: double, precision: 9, scale 4)
- MinElev (type: double, precision: 9, scale 4)
- MaxElev (type: double, precision: 9, scale 4)
- Gradient (type: double, precision: 9, scale 4)

Add the following field to the GeometricDesign layer (use the add field-batch tool):

- Geometric (type: double, precision: 5, scale 2)
- Lat (type: double, precision: 8, scale 5)
- Long (type: double, precision: 8, scale 5)
- COMID (type: long, precision: 9)

Join the nhdflopline layer with the FlowlineAttributesFlow table based on the COMID field.

Use the field calculator within the nhdflopline attribute table, with the appropriate metric to imperial conversion to populate the following fields:

- LENGTHMi (from LENGTHKM – kilometers to miles)

- DrainMia (from CumDrainage – square kilometers to square miles (sq mi))
- MinElev (from MinElevSmo – meters to feet)
- MaxElev (from MaxElevSmo – meters to feet)
- Gradient ((MaxElev-MinElev)/LENGTHMI).

Unjoin the FlowlineAttributesFlow table.

Label the “nhdfowline” layer based new “LengthMi” field – note: this field shows the cumulative drainage at the *end* of the line segment, which is rarely more than 2-3 miles in between nodes.

Calculate the geometric break points (i.e., for a 500 sq mi watershed: 500, 250, 125, 62.5, 31, 15, 7, 4, 2).

It is recommended to change the symbology (Symbology: Show Quantities: Classification (Manual)) of the actual flowline to reflect the drainage. This will help identify when and where sites need to be allocated.

Start a new editing session, with the GeometricDesign layer as your target layer.

Add a new point within this layer to the pour point for the watershed (500 sq mi in this case).

Travel upstream through the mainstem and “find” the next place on the stream where the river drainage brackets 250 sq mi. Use the catchment shapefile layer to identify more precisely the drainage value if needed.

Populate the “Geometric” field within the GeometricDesign layer accordingly to the identified drainage level, then change the symbology (Symbology: Categories: Unique Values: Geometric field) of this layer to reflect the drainage levels.

Proceed through the watershed (either around the outer portions or start with largest values and work in), adding points accordingly to each geometric level. Change the symbology to find areas or levels that were missed. Note – the drainage level must be exact. Use the catchment shapefile to subtract drainage areas from larger drainage areas until the exact drainage level is reached. It is ok to “skip” a geometric level if it is not exactly reached. Sometimes there are large tributaries whose contribution to the mainstem skips a drainage level.

Populate the COMID (manually), and Lat/Long (right click on field and select calculate geometry – lat = x-coordinates and long = y-coordinates) accordingly for reference within the GeometricDesign Layer

Once sites are selected in this fashion, they will need to be snapped to a bridge or access point.

Additional sites should be placed at pour points of subwatersheds (12-digit HUCs) to meet TMDL document requirements.

Once the initial sites are selected, the following features are taken into account to move or add sites:

- Permitted facilities
- Urban areas
- Historical sampling sites
- Assessment Unit IDs (AUID)
- External stakeholder information
- Resources - maximum of 35 sites per project

After refining site selections, there may be additional sites added to ensure spatial representation of the project area.

Sites may be removed or changed after site reconnaissance if there are problems accessing the site or if sites are dry.

Notes regarding the NHD dataset:

All units are initially set to metric and need to be converted to imperial.

Within the nhdfLOWline layer, the GNIS\_Name/ID refers to the whole river name and ID, while the COMID is a unique identifier for the particular segment.

There is *not* a value GNIS\_Name/ID for every river, especially where primary streams and ditches are concerned.

Segments within the nhdfLOWline layer are based on linear miles between “nodes,” which are broken up (typically) by tributary. Typically these lengths are less than 2-3 miles.

The cumulative drainage values in the NHD dataset have been compared against other and deemed “reasonable” (read – not statistically compared). Also note that the drainage is calculated through the model to be at the pour point of that segment.

The elevation values, however, are **not** reliable and require supervision. These values are calculated from the associated digital elevation model (DEM) and sometimes have null values for either the maximum or minimum elevation values. In addition, the length of the stream is not long enough (i.e. >1 mile) to calculate gradient. In either case, this associated value is helpful to identify contour changes against a USGS contour map. However, to note the calculated gradient from the NHD information has been observed to be within several tenths of mile compared to a manual calculation of gradient.


Important tables from NHD

- FlowlineAttributesFlow (found in: Region 05, Version 01\_02, Catchment Flowline Attributes)
  - Key fields: CumDrainag, Max ElevRaw, MinElevSmo,

Important Layers from NHD

- Region 05, Version 01\_01, Catchment Shapefile
- Region 05, Version 01\_02, National Hydrography Dataset

## Attachment 2. IDEM Site Reconnaissance Form.

	<b>Site Reconnaissance Form</b>		<b>EPA Site Identifier</b>	<b>Rank</b>
			Recon #: _____ Trip #: _____	

Site Number: _____	Stream: _____	County: _____	
--------------------	---------------	---------------	--

Location Description: \_\_\_\_\_

Reconnaissance Data Collected				Landowner/Contact Information		
Recon Date		Crew Members		First Name		Last Name
Avg. Width (m)	Avg. Depth (m)	Max. Depth (m)	Nearest Town			Street Address
Water Present? <input type="checkbox"/>	Site Wadeable? <input type="checkbox"/>	Riffle/Run Present? <input type="checkbox"/>	Road/Public Access Possible? <input type="checkbox"/>			City
Site Impacted by Livestock? <input type="checkbox"/>	Collect Sediment? <input type="checkbox"/>	Gauge Present? <input type="checkbox"/>				State      Zip
						Telephone      E-Mail Address
			Pamphlet Distributed? <input type="checkbox"/>	Please Call In Advance? <input type="checkbox"/>	Results Requested? <input type="checkbox"/>	

Rating, Results, Comments, and Planning			
<b>Site Rating By Category (1=easy, 10=difficult)</b>  <div style="border: 1px solid black; padding: 2px; margin-bottom: 5px;">Access Route</div> <div style="border: 1px solid black; padding: 2px; margin-bottom: 5px;">Safety Factor</div> <div style="border: 1px solid black; padding: 2px;">Sampling Effort</div>	<div style="border: 1px solid black; padding: 2px;"> <b>Reconnaissance Decision</b>             Pre-Recon            Recon In process            Approved Site            No, Landowner denied access            No, Dry            No, Stream channel missing            No, Physical barriers            No, Impounded stream            No, Marsh/Wetland            No, Bridge gone or not accessible            No, Unsafe due to traffic or location            No, Site impacted by backwater            No, Other         </div>	<b>Equipment Selected</b>  <div style="border: 1px solid black; height: 100px;"></div>	<b>Circle Equipment Needed</b>  Backpack Boat Tote/barge Longline Scooter Seine Weighted Handline Waders Gill Net

**Comments**

**Sketch of Stream & Access Route – Indicate Flow, Direction, Obstacles, & Land Use (Use Back of Page, if Necessary)**



### Attachment 3: Blank Stream Sampling Field Data Sheet

<b style="font-size: 1.2em; margin-left: 10px;">Stream Sampling Field Data Sheet</b>										Analysis Set #	EPA Site ID	Rank
Sample #	Site #		Sample Medium				Sample Type		Duplicate Sample #			
Stream Name:						River Mile:		County:				
Site Description:												
Survey Crew Chief	Sample Collectors				Sample Collected		HydroLab #	Water Depth/Gage Ht (ft)	Water Flow (cfs/sec)	Flow Estimated?	Algae?	Aquatic Life?
	1	2	3	4	Date	Time						
Sample Taken?			Aliquots			Water Flow Type			Water Appearance			Canopy Closed %
<input type="checkbox"/> Yes <input type="checkbox"/> No; Frozen <input type="checkbox"/> No; Stream Dry <input type="checkbox"/> No; Other <input type="checkbox"/> No; Owner refused Access			<input type="checkbox"/> 1 <input type="checkbox"/> 2 <input type="checkbox"/> 3 <input type="checkbox"/> 4 <input type="checkbox"/> 8 <input type="checkbox"/> 8 <input type="checkbox"/> 12 <input type="checkbox"/> 24 <input type="checkbox"/> 48 <input type="checkbox"/> 72 <input type="checkbox"/> AS-Flow			<input type="checkbox"/> Riffle <input type="checkbox"/> Dry <input type="checkbox"/> Pool <input type="checkbox"/> Run <input type="checkbox"/> Flood <input type="checkbox"/> Glide <input type="checkbox"/> Eddy <input type="checkbox"/> Other			<input type="checkbox"/> Clear <input type="checkbox"/> Green <input type="checkbox"/> Shown <input type="checkbox"/> Murky <input type="checkbox"/> Black <input type="checkbox"/> Other <input type="checkbox"/> Brown <input type="checkbox"/> Gray (Septic/Sewage)			<input type="checkbox"/> 0-20% <input type="checkbox"/> 80-85% <input type="checkbox"/> 20-40% <input type="checkbox"/> 80-100% <input type="checkbox"/> 40-80%
Special Notes:												

**Field Data:**

Date (m/d/yy)	24-hr Time (hh:mm)	D.O. (mg/l)	pH	Water Temp (°C)	Spec Cond (µohms/cm)	Turbidity (NTU)	% Sat.	Chlorine (mg/l)	Chloride (mg/l)	Chlorophyll (mg/l)	Weather Codes		
SC	WD	WS	AT										

Measurement Flags	<	< Min. Meter Measurement	Weather Code Definitions			
	>	> Max. Meter Measurement				
E		Estimated (See Comments)	SC	WD	WS	AT
R		Rejected (See Comments)	Sky Conditions	Wind Direction	Wind Strength	Air Temp
			1 Clear	8 Rain	00 North (0 degrees)	0 Calm
			2 Scattered	9 Snow	09 East (90 degrees)	1 Light
			3 Partly	10 Sleet	18 South (180 degrees)	2 Mod./Light
			4 Cloudy		27 West (270 degrees)	3 Moderate
			5 Mist			4 Mod./Strong
			6 Fog			5 Strong
			7 Shower			6 Gale
						1 < 32
						2 33-45
						3 46-60
						4 61-75
						5 76-85
						6 > 86

**Field Calibrations:**

Date (m/d/yy)	Time (hh:mm)	Calibrator Initials	Calibrations			
			Type	Meter #	Value	Units

Calibration Type	pH	DO	Turbidity

**Preservatives/Bottle Lots:**

Group: Preservative	Preservative Lot #	Bottle Type	Bottle Lot #

Groups: Preservatives		Bottle Types	
GC General Chemistry: Ice	2000P	2000mL Plastic, Narrow Mouth	
Nx Nutrients: H2SO4	1000P	1000mL Plastic, Narrow Mouth	
Metals: HNO3	500P	500mL Plastic, Narrow Mouth	
CN Cyanide: NaOH	250P	250mL Plastic, Narrow Mouth	
Oil & Grease: H2SO4	1000G	1000mL Glass, Narrow Mouth	
Toxics: Ice	500G	500mL Glass, Wide Mouth	
Ecol: Bacteriology: Ice	250G	250mL Glass, Wide Mouth	
VOA Volatile Organics: HCl & Thiosulfate	125G	125mL Glass, Wide Mouth	
Pest Pesticides: Ice	40GV	40mL Glass Vial	
Phen Phenols: H2SO4	120PB	120mL Plastic (Bacteria Only)	
Sed Sediment: Ice	1000PF	1000mL Plastic, Coming Filter	
Gly Glyphosate: Thiosulfate	500PF	500mL Plastic, Coming Filter	
Hg Mercury(1631): HCl	50P	50mL Plastic	
Cr6 Chromium(VI)(1636): NaOH	250T	250mL Teflon	
MeHg Methyl Mercury(1630): HCl	500T	500mL Teflon	
	125T	125mL Teflon	

Data Entered By: \_\_\_\_\_ QC1: \_\_\_\_\_  
 QC2: \_\_\_\_\_

Stream Sampling Field Data Sheet

## Attachment 4: Fish Collection Data Sheet

IDEM  
 OWQ-WATERSHED ASSESSMENT AND PLANNING BRANCH


Event ID \_\_\_\_\_ Voucher jars \_\_\_\_\_ Unknown jars \_\_\_\_\_ Equipment \_\_\_\_\_ Page \_\_\_\_\_ of \_\_\_\_\_  
 Voltage \_\_\_\_\_ Time fished (sec) \_\_\_\_\_ Distance fished (m) \_\_\_\_\_ Max. depth (m) \_\_\_\_\_ Avg. depth (m) \_\_\_\_\_  
 Avg. width (m) \_\_\_\_\_ Bridge in reach \_\_\_\_\_ Is reach representative \_\_\_\_\_ If no, why \_\_\_\_\_  
 Elapsed time at site (hh:mm) \_\_\_\_\_; \_\_\_\_\_ Comments \_\_\_\_\_

Museum data: Initials \_\_\_\_\_ ID date \_\_\_\_\_ Jar count \_\_\_\_\_ Fish Total \_\_\_\_\_

Coding for Anomalies: D – deformities E – eroded fins L – lesions T – tumor M – multiple DELT anomalies O – other (A – anchor worm C – leeches  
 W – swirled scales Y – popeye S – emaciated F – fungus P – parasites) H – heavy L – light (these codes may be combined with above codes)

TOTAL # OF FISH				WEIGHT (s)			ANOMALIES						
				(mass g)			(length mm)						
							Min length	D	E	L	T	M	O
							Max length						
V		P											
							Min length	D	E	L	T	M	O
							Max length						
V		P											
							Min length	D	E	L	T	M	O
							Max length						
V		P											
							Min length	D	E	L	T	M	O
							Max length						
V		P											
							Min length	D	E	L	T	M	O
							Max length						
V		P											
							Min length	D	E	L	T	M	O
							Max length						
V		P											

## Attachment 5: Macroinvertebrate Header Form.



### Office of Water Quality: Macroinvertebrate Header

L-Site #	Event ID	Stream Name	Location	County	Surveyor

Sample Date	Sample #	Macro#	# Containers

**Macro Sample Type:**

☐ Black Light  
☐ CPOM  
☐ Hester-Dendy

☐ Kick  
☐ MHAB  
☐ Qualitative

☐ Normal  
☐ Duplicate  
☐ Replicate

☐ Habitat Complete    ☐ Sample Quality Rejected

### Riparian Zone/Instream Features

**Watershed Erosion:**

☐ Heavy  
☐ Moderate  
☐ None

**Watershed NPS Pollution:**

☐ No Evidence  
☐ Obvious Sources  
☐ Some Potential Sources

**Stream Depth Riffle (m):**

**Stream Depth Run (m):**

**Stream Depth Pool (m):**

**Distances Riffle-Riffle (m):**

**Distances Bend-Bend (m):**

**Stream Width (m):**

**High Water Mark (m):**

**Velocity (ft/s):**

**Stream Type:**

☐ Cold  
☐ Warm

**Turbidity (Est):**

☐ Clear    ☐ Slightly Turbid  
☐ Opaque    ☐ Turbid

**Salinity (mg/L):**

**ORP (mV):**

☐ Channelization    ☐ Dam Present

**Predominant Surrounding Land Use:** ☐ Forest ☐ Field/Pasture ☐ Agricultural ☐ Residential ☐ Commercial ☐ Industrial

Other

### Sediment

**Sediment Odors:** ☐ Normal ☐ Sewage ☐ Petroleum ☐ Chemical ☐ Anaerobic ☐ None Other

**Sediment Deposits:** ☐ Sludge ☐ Sawdust ☐ Paper Fiber ☐ Sand ☐ Relic Shells Other

**Sediment Oils:** ☐ Absent ☐ Moderate ☐ Profuse ☐ Slight

☐ Are the undersides of stones, which are not deeply embedded, black?

### Substrate Components

(Note: Select from 0%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, or 100% for each inorganic/ organic substrate component)

Inorganic Substrate Components (% Diameter)							Organic Substrate Components (% Type)			
Bedrock	Boulder (>10 in)	Cobble (2.5-10 in)	Gravel (0.1-2.5 in)	Sand (gritty)	Silt	Clay (slick)	Detritus (sticks, wood)	Detritus (CPOM)	Muck/Mud (black, fine FPOM)	Marl(gray w/ shell fragments)

### Water Quality

**Water Odors:** ☐ Normal ☐ Sewage ☐ Petroleum ☐ Chemical ☐ None Other

**Water Surface Oils:** ☐ Slick ☐ Sheen ☐ Glob ☐ Flocks ☐ None

## Attachment 6: Biological Samples Field Chain-of-custody Form

[illegible]

**Attachment 7: Blank OWQ Biological Studies QHEI (Qualitative Habitat Evaluation Index) form  
(front)**



# OWQ Biological QHEI (Qualitative Habitat Evaluation Index)

Sample #	bioSample #	Stream Name	Location
Surveyor	Sample Date	County	Macro Sample Type
<input type="checkbox"/> Habitat Complete			QHEI Score: <input type="text"/>

1] **SUBSTRATE** Check ONLY Two predominant substrate TYPE BOXES; estimate % and check every type present

BEST TYPES		OTHER TYPES		ORIGIN		QUALITY	
PREDOMINANT	PRESENT TOTAL %	PREDOMINANT	PRESENT TOTAL %				
P/G R/R		P/G R/R		<input type="checkbox"/> LIMESTONE [1]	<input type="checkbox"/> HEAVY [-2]	Substrate	<input type="text"/>
<input type="checkbox"/> BLDR/SLABS [10]	<input type="checkbox"/>	<input type="checkbox"/> HARDPAN [4]	<input type="checkbox"/>	<input type="checkbox"/> TILLS [1]	<input type="checkbox"/> MODERATE [-1]		
<input type="checkbox"/> BOULDER [9]	<input type="checkbox"/>	<input type="checkbox"/> DETRITUS [3]	<input type="checkbox"/>	<input type="checkbox"/> WETLANDS [0]	<input type="checkbox"/> NORMAL [0]		
<input type="checkbox"/> COBBLE [8]	<input type="checkbox"/>	<input type="checkbox"/> MUCK [2]	<input type="checkbox"/>	<input type="checkbox"/> HARDPAN [0]	<input type="checkbox"/> FREE [1]		
<input type="checkbox"/> GRAVEL [7]	<input type="checkbox"/>	<input type="checkbox"/> SILT [2]	<input type="checkbox"/>	<input type="checkbox"/> SANDSTONE [0]			
<input type="checkbox"/> SAND [6]	<input type="checkbox"/>	<input type="checkbox"/> ARTIFICIAL [0]	<input type="checkbox"/>	<input type="checkbox"/> RIP/RAP [0]	<input type="checkbox"/> EXTENSIVE [-2]	Maximum	<input type="text"/>
<input type="checkbox"/> BEDROCK [5]	<input type="checkbox"/>	(Score natural substrates; ignore sludge from point-sources)		<input type="checkbox"/> LACUSTRINE [0]	<input type="checkbox"/> MODERATE [-1]		
NUMBER OF BEST TYPES: <input type="checkbox"/> 4 or more [2] <input type="checkbox"/> 3 or less [0]				<input type="checkbox"/> SHALE [-1]	<input type="checkbox"/> NORMAL [0]		
				<input type="checkbox"/> COAL FINES [-2]	<input type="checkbox"/> NONE [1]		

Comments

2] **INSTREAM COVER** Indicate presence 0 to 3 and estimate percent: 0-Absent; 1-Very small amounts or if more common of marginal quality; 2-Moderate amounts, but not of highest quality or in small amounts of highest quality; 3-Highest quality in moderate or greater amounts (e.g., very large boulders in deep or fast water, large diameter log that is stable, well developed root wad in deep/fast water, or deep, well-defined, functional pools.)

% Amount		% Amount		% Amount		AMOUNT	
<input type="checkbox"/> UNDERCUT BANKS [1]	<input type="checkbox"/>	<input type="checkbox"/> POOLS > 70cm [2]	<input type="checkbox"/>	<input type="checkbox"/> OXBOWS, BACKWATERS [1]	<input type="checkbox"/>	<input type="checkbox"/> EXTENSIVE > 75% [11]	Cover
<input type="checkbox"/> OVERHANGING VEGETATION [1]	<input type="checkbox"/>	<input type="checkbox"/> ROOTWADS [1]	<input type="checkbox"/>	<input type="checkbox"/> AQUATIC MACROPHYTES [1]	<input type="checkbox"/>	<input type="checkbox"/> MODERATE 25 - 75% [7]	
<input type="checkbox"/> SHALLOWS (IN SLOW WATER) [1]	<input type="checkbox"/>	<input type="checkbox"/> BOULDERS [1]	<input type="checkbox"/>	<input type="checkbox"/> LOGS OR WOODY DEBRIS [1]	<input type="checkbox"/>	<input type="checkbox"/> SPARSE 5 - < 25% [3]	
<input type="checkbox"/> ROOTMATS [1]	<input type="checkbox"/>					<input type="checkbox"/> NEARLY ABSENT < 5% [1]	
						Maximum	
							<input type="text"/>

Comments

3] **CHANNEL MORPHOLOGY** Check ONE in each category (Or 2 & average)

SINUOSITY		DEVELOPMENT		CHANNELIZATION		STABILITY		
<input type="checkbox"/> HIGH [4]	<input type="checkbox"/>	<input type="checkbox"/> EXCELLENT [7]	<input type="checkbox"/>	<input type="checkbox"/> NONE [6]	<input type="checkbox"/>	<input type="checkbox"/> HIGH [3]	Channel	
<input type="checkbox"/> MODERATE [3]	<input type="checkbox"/>	<input type="checkbox"/> GOOD [5]	<input type="checkbox"/>	<input type="checkbox"/> RECOVERED [4]	<input type="checkbox"/>	<input type="checkbox"/> MODERATE [2]		
<input type="checkbox"/> LOW [2]	<input type="checkbox"/>	<input type="checkbox"/> FAIR [3]	<input type="checkbox"/>	<input type="checkbox"/> RECOVERING [3]	<input type="checkbox"/>	<input type="checkbox"/> LOW [1]		
<input type="checkbox"/> NONE [1]	<input type="checkbox"/>	<input type="checkbox"/> POOR [1]	<input type="checkbox"/>	<input type="checkbox"/> RECENT OR NO RECOVERY [1]	<input type="checkbox"/>			
							Maximum	<input type="text"/>

Comments

4] **BANK EROSION AND RIPARIAN ZONE** Check ONE in each category for EACH BANK (Or 2 per bank & average)

River right looking downstream		RIPARIAN WIDTH		FLOOD PLAIN QUALITY		CONSERVATION TILLAGE		
L R		L R		L R		L R		
<input type="checkbox"/> NONE/LITTLE [3]	<input type="checkbox"/>	<input type="checkbox"/> WIDE > 50m [4]	<input type="checkbox"/>	<input type="checkbox"/> FOREST, SWAMP [3]	<input type="checkbox"/>	<input type="checkbox"/> URBAN OR INDUSTRIAL [0]	Riparian	
<input type="checkbox"/> MODERATE [2]	<input type="checkbox"/>	<input type="checkbox"/> MODERATE 10-50m [3]	<input type="checkbox"/>	<input type="checkbox"/> SHRUB OR OLD FIELD [2]	<input type="checkbox"/>	<input type="checkbox"/> MINING / CONSTRUCTION [0]		
<input type="checkbox"/> HEAVY/SEVERE [1]	<input type="checkbox"/>	<input type="checkbox"/> NARROW 5-10m [2]	<input type="checkbox"/>	<input type="checkbox"/> RESIDENTIAL, PARK, NEW FIELD [1]	<input type="checkbox"/>			
		<input type="checkbox"/> VERY NARROW [1]	<input type="checkbox"/>	<input type="checkbox"/> FENCED PASTURE [1]	<input type="checkbox"/>			
		<input type="checkbox"/> NONE [0]	<input type="checkbox"/>	<input type="checkbox"/> OPEN PASTURE, ROW CROP [0]	<input type="checkbox"/>	Indicate predominant land use(s) past 100m riparian.		
							Maximum	<input type="text"/>

Comments

5] **POOL/GLIDE AND RIFFLE/RUN QUALITY**

MAXIMUM DEPTH		CHANNEL WIDTH		CURRENT VELOCITY		Recreation Potential		
Check ONE (ONLY!)		Check ONE (Or 2 & average)		Check ALL that apply		(Circle one and comment on back)		
<input type="checkbox"/> > 1m [6]	<input type="checkbox"/>	<input type="checkbox"/> POOL WIDTH > RIFFLE WIDTH [2]	<input type="checkbox"/>	<input type="checkbox"/> TORRENTIAL [-1]	<input type="checkbox"/>	<input type="checkbox"/> SLOW [1]	Pool/Current	
<input type="checkbox"/> 0.7 - < 1m [4]	<input type="checkbox"/>	<input type="checkbox"/> POOL WIDTH = RIFFLE WIDTH [1]	<input type="checkbox"/>	<input type="checkbox"/> VERY FAST [1]	<input type="checkbox"/>	<input type="checkbox"/> INTERSTITIAL [-1]		
<input type="checkbox"/> 0.4 - < 0.7m [2]	<input type="checkbox"/>	<input type="checkbox"/> POOL WIDTH < RIFFLE WIDTH [0]	<input type="checkbox"/>	<input type="checkbox"/> FAST [1]	<input type="checkbox"/>	<input type="checkbox"/> INTERMITTENT [-2]		
<input type="checkbox"/> 0.2 - < 0.4m [1]	<input type="checkbox"/>		<input type="checkbox"/>	<input type="checkbox"/> MODERATE [1]	<input type="checkbox"/>	<input type="checkbox"/> EDDIES [1]		
<input type="checkbox"/> < 0.2m [0] [metric = 0]	<input type="checkbox"/>		<input type="checkbox"/>	Indicate for reach - pools and riffles.				
							Maximum	<input type="text"/>

Comments

Indicate for functional riffles; Best areas must be large enough to support a population of riffle-obligate species:


RIFFLE DEPTH		RUN DEPTH		RIFFLE/RUN SUBSTRATE		RIFFLE/RUN EMBEDDEDNESS		
Check ONE (ONLY!)		Check ONE (Or 2 & average)		Check ONE (Or 2 & average)		Check ONE (Or 2 & average)		
<input type="checkbox"/> BEST AREAS > 10cm [2]	<input type="checkbox"/>	<input type="checkbox"/> MAXIMUM > 50cm [2]	<input type="checkbox"/>	<input type="checkbox"/> STABLE (e.g., Cobble, Boulder) [2]	<input type="checkbox"/>	<input type="checkbox"/> NONE [2]	Riffle/Run	
<input type="checkbox"/> BEST AREAS 5 - 10cm [1]	<input type="checkbox"/>	<input type="checkbox"/> MAXIMUM < 50cm [1]	<input type="checkbox"/>	<input type="checkbox"/> MOD. STABLE (e.g., Large Gravel) [1]	<input type="checkbox"/>	<input type="checkbox"/> LOW [1]		
<input type="checkbox"/> BEST AREAS < 5cm [metric = 0]	<input type="checkbox"/>		<input type="checkbox"/>	<input type="checkbox"/> UNSTABLE (e.g., Fine Gravel, Sand) [0]	<input type="checkbox"/>	<input type="checkbox"/> MODERATE [0]		
						<input type="checkbox"/> EXTENSIVE [-1]		
							Maximum	<input type="text"/>

Comments

6] <b>GRADIENT</b> ( ft/ mi )		7] <b>GRADIENT</b> ( mi <sup>2</sup> )		% POOL		% GLIDE	
<input type="checkbox"/> VERY LOW - LOW [2-4]	<input type="checkbox"/>	<input type="checkbox"/> VERY LOW - LOW [2-4]	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	Gradient
<input type="checkbox"/> MODERATE [6-10]	<input type="checkbox"/>	<input type="checkbox"/> MODERATE [6-10]	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	
<input type="checkbox"/> HIGH - VERY HIGH [10-6]	<input type="checkbox"/>	<input type="checkbox"/> HIGH - VERY HIGH [10-6]	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	Maximum
							<input type="text"/>

IDEM 11/15/12

## Attachment 7 (continued). IDEM OWQ Biological QHEI (back).



**OWQ Biological QHEI (Qualitative Habitat Evaluation Index)**

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COMMENT \_\_\_\_\_

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<b>A-CANOPY</b> <input type="checkbox"/> > 85% - Open <input type="checkbox"/> 55% - < 85% <input type="checkbox"/> 30% - < 55% <input type="checkbox"/> 10% - < 30% <input type="checkbox"/> < 10% - Closed	<b>B-AESTHETICS</b> <input type="checkbox"/> Nuisance algae <input type="checkbox"/> Invasive macrophytes <input type="checkbox"/> Excess turbidity <input type="checkbox"/> Discoloration <input type="checkbox"/> Foam/Scum	<input type="checkbox"/> Oil sheen <input type="checkbox"/> Trash/Litter <input type="checkbox"/> Nuisance odor <input type="checkbox"/> Sludge deposits <input type="checkbox"/> CSOs/SSOs/Outfalls	<b>C-RECREATION</b> Area      Depth Pool <input type="checkbox"/> > 100 ft <sup>2</sup> <input type="checkbox"/> > 3 ft	<b>D-MAINTENANCE</b> <input type="checkbox"/> Public <input type="checkbox"/> Private <input type="checkbox"/> Active <input type="checkbox"/> Historic Successions: <input type="checkbox"/> Young <input type="checkbox"/> Old <input type="checkbox"/> Spray <input type="checkbox"/> Islands <input type="checkbox"/> Scoured Snag: <input type="checkbox"/> Removed <input type="checkbox"/> Modified Leveed: <input type="checkbox"/> One sided <input type="checkbox"/> Both banks <input type="checkbox"/> Relocated <input type="checkbox"/> Cutoffs Bedload: <input type="checkbox"/> Moving <input type="checkbox"/> Stable <input type="checkbox"/> Armoured <input type="checkbox"/> Skumps <input type="checkbox"/> Impounded <input type="checkbox"/> Desiccated <input type="checkbox"/> Flood control <input type="checkbox"/> Drainage	<b>E-ISSUES</b> <input type="checkbox"/> WWTP <input type="checkbox"/> CSO <input type="checkbox"/> NPDES <input type="checkbox"/> Industry <input type="checkbox"/> Urban <input type="checkbox"/> Hardened <input type="checkbox"/> Dirt & Grime <input type="checkbox"/> Contaminated <input type="checkbox"/> Landfill BMPs: <input type="checkbox"/> Construction <input type="checkbox"/> Sediment <input type="checkbox"/> Logging <input type="checkbox"/> Irrigation <input type="checkbox"/> Cooling Erosion: <input type="checkbox"/> Bank <input type="checkbox"/> Surface <input type="checkbox"/> False bank <input type="checkbox"/> Manure <input type="checkbox"/> Lagoon <input type="checkbox"/> Wash H <sub>2</sub> O <input type="checkbox"/> Tile <input type="checkbox"/> H <sub>2</sub> O Table Mine: <input type="checkbox"/> Acid <input type="checkbox"/> Quarry Flow: <input type="checkbox"/> Natural <input type="checkbox"/> Stagnant <input type="checkbox"/> Wetland <input type="checkbox"/> Park <input type="checkbox"/> Golf <input type="checkbox"/> Lawn <input type="checkbox"/> Home <input type="checkbox"/> Atmospheric deposition <input type="checkbox"/> Agriculture <input type="checkbox"/> Livestock
---	--	--	---	---	--

Looking upstream (> 10m, 3 readings ≤ 10m, 1 reading in middle); Round to the nearest whole percent

	Right %	Middle %	Left %	Total Average %
% open	X	X	X	

---

Stream Drawing: \_\_\_\_\_







## Attachment 9: Sample Analysis Request form.



Indiana Department of Environmental Management  
Office of Water Quality  
Watershed Planning and Assessment Branch  
[www.idem.in.gov](http://www.idem.in.gov)

### Water Sample Analysis Request

Project Name: \_\_\_\_\_ Composite ☐ Grab ☒

OWQ Sample Set	1	IDEM Sample Nos.	
Crew Chief		Lab Sample Nos.	
Collection Date		Lab Delivery Date	

#### Anions and Physical Parameters

Parameter	Test Method	Total	Dissolved
Alkalinity (as CaCO <sub>3</sub> )	EPA 310.2	<input checked="" type="checkbox"/> **	<input type="checkbox"/>
Total Solids	SM 2540B	<input checked="" type="checkbox"/> **	
Suspended Solids	SM 2540D	<input checked="" type="checkbox"/> **	
Dissolved Solids	SM 2540C		<input checked="" type="checkbox"/> **
Sulfate	EPA 375.2	<input checked="" type="checkbox"/> **	<input type="checkbox"/> **
Chloride	SM 4500Cl-E	<input checked="" type="checkbox"/> **	<input type="checkbox"/>
Hardness (as CaCO <sub>3</sub> )	EPA 130.1	<input checked="" type="checkbox"/> **	<input type="checkbox"/>
Fluoride	380-75WE	<input type="checkbox"/> **	<input type="checkbox"/>
Silica (Reactive)	SM 4500-SiD	<input type="checkbox"/> **	<input type="checkbox"/>

#### Priority Pollutant Metals Water Parameters

Parameter	Test Method	Total	Dissolved
Antimony	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Arsenic	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Beryllium	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Cadmium	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Chromium (Hex)	SM 3500Cr-D	<input type="checkbox"/>	<input type="checkbox"/>
Chromium (Total)	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Copper	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Lead	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Mercury	EPA 245.1	<input type="checkbox"/>	<input type="checkbox"/>
Nickel	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Selenium	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Silver	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Thallium	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Zinc	200.7	<input type="checkbox"/>	<input type="checkbox"/>

#### Cations and Secondary Metals Parameters

Parameter	Test Method	Total	Dissolved
Aluminum	200.7, 200.8	<input type="checkbox"/>	<input type="checkbox"/>
Barium	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Boron	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Calcium	200.7, 200.8	<input checked="" type="checkbox"/> ***	<input type="checkbox"/>
Calcium (as CaCO <sub>3</sub> )	SM 3500Ca-D	<input type="checkbox"/>	<input type="checkbox"/>
Cobalt	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Iron	200.7	<input type="checkbox"/>	<input type="checkbox"/>
Magnesium	200.7, 200.8	<input checked="" type="checkbox"/> ***	<input type="checkbox"/>
Manganese	200.8	<input type="checkbox"/>	<input type="checkbox"/>
Potassium	SM 3500-K-D	<input type="checkbox"/>	<input type="checkbox"/>
Sodium	200.7	<input type="checkbox"/>	<input type="checkbox"/>
Strontium	200.7	<input type="checkbox"/>	<input type="checkbox"/>

Send reports (Fed. Ex. or UPS) to: Deliver reports to:

David Jordan - IDEM  
Mail Code 65-40-2 (Shadeland)  
100 N. Senate Ave.  
Indianapolis, IN 46204-2251

David Jordan - IDEM  
STE 100  
2525 North Shadeland Ave.  
Indianapolis, IN 46219  
DJordan@idem.in.gov

#### Organic Water Parameters

Parameter	Test Method	Total
Priority Pollutants: Oranochlorine Pesticides and PCBs	EPA 608	<input checked="" type="checkbox"/>
Polynuclear Aromatic Hydrocarbons	EPA 610	<input type="checkbox"/>
Priority Pollutants: VOCs - Purgeable Organics	EPA 624	<input type="checkbox"/>
Priority Pollutants: Base/Neutral Extractables	EPA 625	<input type="checkbox"/>
Priority Pollutants: Acid Extractables	EPA 625	<input type="checkbox"/>
Phenolics, 4AAP	EPA 420.4	<input type="checkbox"/>
Oil and Grease, Total	EPA 1664A	<input type="checkbox"/>
Semi-volatile Organics & Pesticides	EPA 525.2	<input type="checkbox"/>

#### Nutrient & Organic Water Chemistry Parameters

Parameter	Test Method	Total	Dissolved
Ammonia Nitrogen	EPA 350.1	<input checked="" type="checkbox"/>	<input type="checkbox"/>
CBOD <sub>5</sub>	SM 5210B	<input type="checkbox"/>	
CBOD <sub>u</sub>	SM 5210B	<input type="checkbox"/>	
Total Kjeldahl Nitrogen (TKN)	EPA 351.2	<input checked="" type="checkbox"/>	<input type="checkbox"/>
Nitrate + Nitrite	EPA 353.1	<input checked="" type="checkbox"/>	<input type="checkbox"/>
Dissolved Reactive Phosphorus	SM4500-P	<input type="checkbox"/>	<input type="checkbox"/>
Total Phosphorus	EPA 365.1	<input checked="" type="checkbox"/>	<input type="checkbox"/>
TOC	SM 5310B	<input checked="" type="checkbox"/>	<input type="checkbox"/>
COD (Low Level)	SM 5220D	<input type="checkbox"/>	<input type="checkbox"/>
Cyanide (Total)	EPA 335.4	<input type="checkbox"/>	<input type="checkbox"/>
Cyanide (Free)	SM 4500CN-I	<input type="checkbox"/> *	<input type="checkbox"/>
Cyanide (Amenable)	SM 4500CN-G	<input type="checkbox"/> *	<input type="checkbox"/>

#### Bacteriological Water Parameters

Parameter	Test Method	Total	Dissolved
<i>E. coli</i> (ColiBert Method)	SM9223B	<input type="checkbox"/>	

30 day reporting time required.

#### Notes:

\*\* = DO NOT RUN PARAMETER IF SAMPLE IDENTIFIED AS A BLANK ON THE CHAIN OF CUSTODY

\* = RUN ONLY IF TOTAL CYANIDE IS DETECTED

\*\*\* = Report Calcium, Magnesium as Total Hardness components if Hardness is calculated

Testing Laboratory:  
Indiana State Department of Health (ISDH)  
Environmental Laboratory Division  
550 W. 16th Street  
Indianapolis, IN 46202  
Phone: 317-921-5815 (Ray Beebe)

(Rev. 6/2013)